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A pattern of antimicrobial susceptibility and the correlation of cotrimoxazole and fluoroquinolones in *Escherichia coli* resistant uropathogens in Ibn-Sena hospital in Benghazi-Libya.

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Highlights

- **Escherichia coli strains are the most prevalent bacteria in urinary tract infections.**
- **Dissemination of Escherichia coli resistant strains is one of the main causes of treatment failure.**
- **Cross-resistance to newer antibiotics may be developed by the random utilization of antimicrobial therapy.**
- **Extension of resistance to fluoroquinolones from other antibiotics is evident recently.**
- **A proper selection of antibiotic therapy based on antibiotic susceptibility tests is necessary to attenuate the cross-resistance to the newer generations of antibiotics.**

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ABSTRACT

Uncontrollable use of antibiotics to treat uropathogens is one of the major causes of increasing antimicrobial-resistant *Escherichia coli* (*E. coli*) strains. Therefore the choice of antibiotic therapy is needed to be based on their antibiotic susceptibility tests. We investigated the prevalence and antibiotic susceptibility of *E. coli* in uncomplicated urinary tract infection (UTI) in 1011 samples during 8 months period in Ibn-Sena clinic in Benghazi. The *E. coli* was the most isolated uropathogen (64%). The most resistant antibiotic to this uropathogen was amoxicillin (51.6%), followed by cotrimoxazole (47.7%) and the percentage of ciprofloxacin was 26.6%. When the utilization of ciprofloxacin increased lately as an alternative to cotrimoxazole, its resistance increased and we found that this increase was positively correlated with cotrimoxazole. In conclusion, monitoring and investigating antibiotic-sensitivity patterns may help the physician to prepare reliable strategies for better management and control of *E. coli* infections.

1. Introduction

Uncomplicated urinary tract infections (UTI) are highly prevalent in large portions of the population worldwide. It is the second common infection after respiratory infections (Gupta *et al.*, 2001). The *Escherichia coli* (*E. coli*) is one of the most common bacterial pathogens isolated from both communities acquired and nosocomial patients with UTI (Hryniewicz *et al.*, 2001, Dasgupta *et al.*, 2005). It accounts for more than 80% of non-serious UTIs (Kang *et al.*, 2018).

In 2011, the trimethoprim-sulfamethoxazole (cotrimoxazole) is recommended by the Infectious Diseases Society of America (IDSA) as one of the best choice drugs that may be administered if the rate of resistance of bacteria attributed to acute uncomplicated UTIs is not higher than 20% (Gupta *et al.*, 2011). While the resistance to the cotrimoxazole and other first-line choices such as ampicillin nitrofurantoin is becoming increasingly evasive, fluoroquinolones are commonly recommended as alternatives because it is narrow-spectrum antibiotics that showed a higher range of susceptibility in the *E. coli* strains (Gupta *et al.*, 2011).

Unfortunately, the rate of development of multi-drug resistance in many gram-negative bacteria, especially the *E. coli* becomes hugely evident and this is liable for developing resistance to many unrelated antibiotic classes (George and Levy, 1983). Interestingly,

in recent years the resistance to fluoroquinolones such as ciprofloxacin and levofloxacin is disseminated widely (Karlowksy *et al.*, 2006, Hooton, 2003).

Bacteria produce the extended-spectrum beta-lactamase (ESBL) enzymes extended to cause a resistance to the newer beta-lactam antibiotics as well as to many non-beta lactam antimicrobial agents such as the aminoglycosides and cotrimoxazole (Paterson *et al.*, 2000). A cross-resistance between ciprofloxacin and other non-fluoroquinolones has increased in recent years (Ozyurt *et al.*, 2008).

The ability of the *E. coli* to cause UTI and the trouble in the management of these infections can lead to proliferation of virulent *E. coli* and increase morbidity. Therefore, the health workers must update their knowledge of the current drug resistance. Antibiotic resistance may be changed according to geographic areas because it is directly related to the misuse of antibiotics. This is particularly in a community like Libya where all kinds of antibiotics are available with or without prescription. The objective of this work was to determine the antimicrobial susceptibility of *E. coli* strains isolated from urine samples of patients with UTI collected from Ibn-sena clinic, one of the biggest clinics in Benghazi city. In addition, to find the correlation of resistance between the most commonly two anti-

tibiotics used in the treatment of UTI, the cotrimoxazole and ciprofloxacin. This not only guides general practitioners to prescribe appropriate antibiotics but also for evidence-based recommendations in empirical antibiotic treatment of UTI.

2. Material and methods

2.1. Study design

A retrospective descriptive study at Ibn-Sena clinic in Benghazi-Libya was conducted after approval from the local authority. A total of 1011 samples of patients with urinary tract infections were collected in the period from the 1st of August 2019 to the 30th of April 2020. The samples were of different ages and sex and were collected by early morning samples according to the midstream method and in a sterile container, and sent to the laboratory.

2.2. Bacterial isolates and Antibiotic susceptibility testing

Urine samples were put on cysteine lactose electrolytes deficient (CLED) agar incubated at 37°C for 24 hours. A growth of >105 colony-forming units per mL of any type of organism was considered as significant bacteriuria (Murray et al., 1999). Identification of the *E. coli* was done by observing colonial morphology on CLED medium (Murray et al., 1999). The antibiotic susceptibility of this pathogen was investigated by the Clinical and Laboratory Standards Institute's disc diffusion method (Institute of C. A. L. S., 2006). The following antibiotics were tested: amoxicillin, co-amoxiclav (Augmentin), cotrimoxazole, ciprofloxacin, nitrofurantoin, cefotaxime, ceftriaxone, levofloxacin, gentamicin, and imipenem.

2.3. The relationship of cotrimoxazole and ciprofloxacin resistance

The *E. coli* isolates were grouped as either cotrimoxazole susceptible or resistant. Each group was further subdivided into ciprofloxacin susceptible and resistant organisms. The number of susceptible and resistant organisms in each group was recorded. The statistical significance of cross-resistance between the cotrimoxazole and ciprofloxacin was calculated by Fisher's exact test and Odd ratio was calculated.

3. Result

From a total of one thousand and eleven samples, only 199 samples had shown bacterial growth, while 664 samples had no bacterial growth. The rest of the samples (148) had shown a non-specific growth, Fig. 1. The prevalence of UTI in the eight months was 19.7%. 71.4% of bacterial growth was in the female patients, while 25.6 % of cases were male patients, Fig. 2. The *E. coli* was the most prevalent isolated uropathogen with (64%), followed by *Klebsilla pneumonia* (*K pneumonia*) and *Pseudomonas aeruginosa* (*P. aeruginosa*) (11.6% each). *Proteus mirabilis* (*P. mirabilis*), *Staphylococcus aureus* (*S aureus*), and *Enterococcus faecalis* (*E. faecalis*) had prevalence values of (4.5, 2.5, and 2 % respectively). *Streptococcus pneumonia* (*S. pneumoniae*) and *Actinomyces israelii* (*A. israelii*) had a lower prevalence with a value of (1.5 % each) and *Neisseria gonorrhea* (*N. gonorrhea*) had the lowest rate (0.5%) Fig. 3.

In this study, isolated *E. coli* showed resistance to a wide range of used antimicrobial agents including the recently produced and expensive ones as imipenem, the beta-lactam antimicrobial agent. The most resistant antibiotic to *E. coli* was amoxicillin (51.6%), followed by cotrimoxazole (47.7%), then nitrofurantoin (45.3%). The prevalence of the strain resistance to augmentin was (35.2%). The *E. coli* also showed ciprofloxacin and levofloxacin resistance at a rate of 26.6 & 18 % respectively. The resistance toward gentamicin was lower at 10.9%. 6.3 & 7% of *E. coli* strains were resistant to cefotaxime and ceftriaxone respectively. The resistance to imipenem had the lowest rate of bacterial resistance by 3.9% Fig. 4.

As shown in the present work, 47.5% of cotrimoxazole-resistant *E. coli* were resistant to ciprofloxacin too and 89.4 % of cotrimoxazole susceptible *E. coli* showed susceptibility to ciprofloxacin

antibiotic Fig. 5. The Fisher's exact test at α level of 0.05 revealed that there was a direct relationship between the bacterial resistance of both antibiotic and the Odd ratio was 7.6.

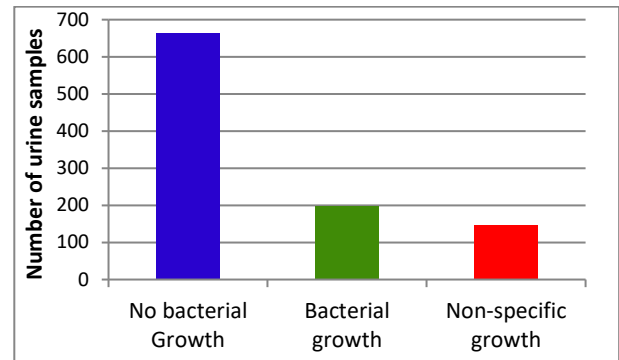


Fig. 1. Results of urine culture tested for bacterial growth in Ibn-sena clinic.

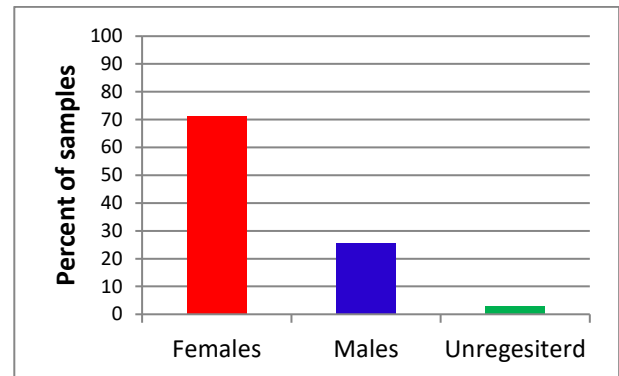


Fig. 2. Gender differences of patients with bacterial growth

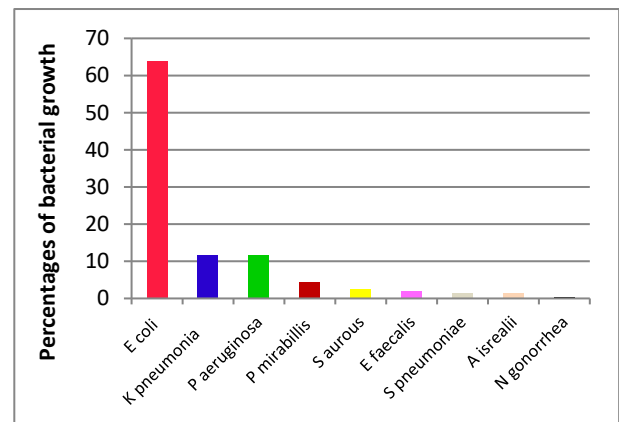


Fig. 3. Prevalence of uropathogens in the collected growth samples.

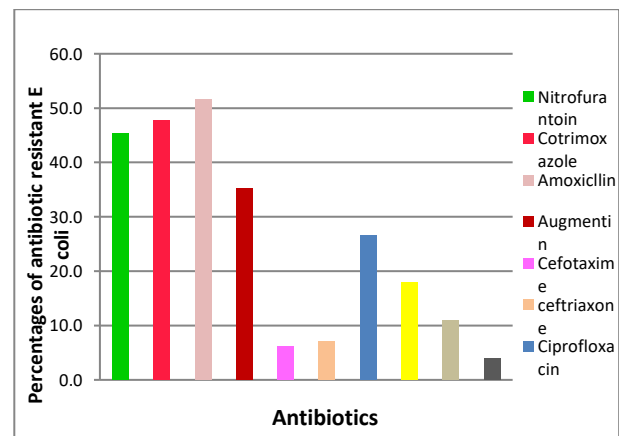


Fig. 4. Prevalence of antibiotics resistance in *E. coli* strains

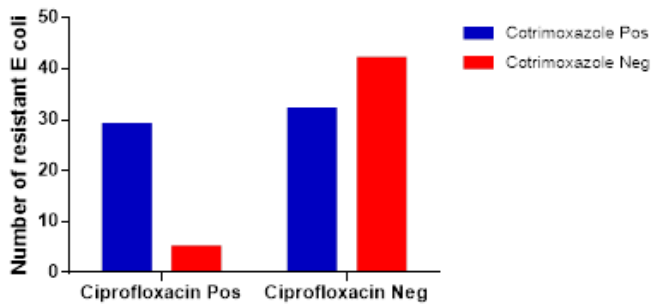


Fig. 5. Cross resistance of cotrimoxazole and ciprofloxacin in *E. coli* strains.

4. Discussion

The progressive increase of antimicrobial resistance has serious consequences such as rapid deterioration of UTI. Therefore, providing information on regional antimicrobial resistance can improve the clinician decision for prescribing the appropriate antibiotic to the specific bacteria and good disease management. In addition, It is an evidence-based recommendation in empirical antibiotic treatment of UTI. Compared to the finding of the present study, there is a variation of the pattern of antimicrobial resistance rate in *E. coli* at different durations in Libya.

The prevalence of *E. coli* in UTI in this study was 64%. A comparable prevalence was reported in other studies (Cullen et al., 2012, Søråas et al., 2013). In the current study, the antibiotic resistance of amoxicillin was (51.6%), followed by cotrimoxazole (47.7%), then nitrofurantoin (45.3%). The prevalence of the bacterial resistance to ciprofloxacin and levofloxacin resistance was at a rate of 26.6 & 18 % respectively. In previous studies, however, the rate of amoxicillin resistant *E. coli* during early 1993 was very high (97%) (Altomi et al., 1997), the trend decreased to 74 and 75 % as shown in two studies were done Tripoli and Benghazi during the early of 2000 (Ghenghesh et al., 2003, Tobgi et al., 2001). A further reduction of amoxicillin resistance in *E. coli* in both cities in 2006-2010 was up to 59 and 57% respectively (Buzayan et al., 2010, Ghenghesh et al., 2009). A better improvement has been noticed in the susceptibility of amoxicillin in this study (48.4%). This may be due to the reduction of amoxicillin prescription in the treatment of UTI recently.

The cotrimoxazole-resistant *E. coli* showed the same pattern of ampicillin resistance. While the rate was higher in different studies in Libya (45-55%) during 1995 (Ghenghesh et al., 2003, Tobgi et al., 2001). It reduced in 2006 to (24-31%) (Buzayan et al., 2010, Ghenghesh et al., 2009). The isolated uropathogens in our study showed increased resistance to cotrimoxazole and this is consistent with other worldwide studies (Ahmed et al., 2000, Hima-Lerible et al., 2003, Ndugulile et al., 2005, Sabrina et al., 2010). Also, Koen B Pouwels and his research group have reported that the prevalence of cotrimoxazole resistance among *E. coli* isolated was high, even after a substantial decline in cotrimoxazole use (Koen et al., 2017).

Conversely, the resistance of isolated uropathogens to ciprofloxacin and levofloxacin had increased. It ranged from 2-17 % in 2002- 2006 in different regions in Libya (Bagar et al., 2007, Buzayan et al., 2010, Ghenghesh et al., 2009). The current study showed an increase in the prevalence of the bacterial resistance to ciprofloxacin and levofloxacin was 26.6 & 18 % respectively. A previous study was done in Benghazi during 2012 where the *E. coli* isolates showed resistance to ciprofloxacin in 40% of samples (Marei et al., 2012). The decrease in susceptibility of *E. coli* to ciprofloxacin was also in line with other studies (Kahlmeter et al., 2015, Seitz et al., 2017, Stapleton et al., 2017). In Germany the rate of resistance raised from 2.2 to 20.2%, in Spain it increased from 14.7 to 30.8%, in the UK, the rate also changed from 0.6 to 15.3% (Kahlmeter et al., 2015). Since the widespread use of these fluoroquinolones as

an alternative to the cotrimoxazole and other first-line therapy in the treatment of UTI causes emerge of their recent resistance.

In the present study, the co-resistance within different classes of antibiotics was significantly high for the cotrimoxazole and ciprofloxacin. ESBL-producing *E. coli* was reported at a high rate of resistance to non-beta lactam antibiotics (Ndugulile et al., 2005, Alhambra et al., 2004). It has been shown that 89.8% of ESBL-producers *E. coli* were non-susceptible to trimethoprim-sulfamethoxazole and 69% were resistant to ciprofloxacin (Dejenie et al., 2019). These findings reported in several studies (Schwaber et al., 2005, Obeng-Nkrumah et al., 2013, Ouedraogo et al., 2016) may indicate that ESBL-producing Enterobacteriaceae were the major cause of resistance to various antibiotics classes. The emergence of resistance in different classes of antimicrobial agents may be explained by the fact that ESBL is plasmid-mediated enzymes that are transferable between one bacterium to another and such transferable plasmids also code for resistance determinants to antimicrobial agents other than beta-lactams (Paterson, 2006).

In conclusion, the results presented in this study indicate that the empirical use of antimicrobial agents, in particular cotrimoxazole and fluoroquinolones, in the treatment of UTI should be reconsidered to develop clear guides to reduce the development of further resistance. Furthermore, periodic antimicrobial resistance surveillance systems should be recommended in different regions and durations to detect the emergence of resistance patterns. This helps in the selection of the most efficacious empirical therapy at the local level.

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References

- Ahmed, A., Osman, H., Mansour, A., Musa, H., Ahmed, A., Z.Karrar & Hassan, H. (2000) 'Antimicrobial agent resistance in bacterial isolates from patients with diarrhea and urinary tract infection in Sudan', *Am J Trop Med Hyg*, 63, pp. 259-263.
- Alhambra, A., Cuadros, J., Cacho, J., Gómez-Garcés, J. & Ji. Alós (2004) 'In vitro susceptibility of recent antibiotic-resistant urinary pathogens to ertapenem and 12 other antibiotics', *J Anti-microb Chemother*, 53, pp. 1090-1094.
- Altomi, A., Abeid, S., Tloba, S., Rhouma, A., Daw, M. & Ghenghesh, K. (1997) Urinary tract infections in Tripoli and drug sensitivity of the causative agents. T. The First Conference on Postgraduate Studies in Medical Sciences, Benghazi, Libya.
- Bagar, S., Hussein, A., Elahwel, A. & Alani, S. (2007) 'Antibiotic resistance pattern of urinary tract isolates', *Jamahiriya Med J*, 7, pp. 116-121.
- Buzayan, M., Tobgi, R. & Taher, I. (2010) 'Detection of extended-spectrum β -lactamases among urinary *Escherichia coli* and *Klebsiella pneumoniae* from two centers', *Jamahiriya Med J*, 10, pp. 6-10.
- Cullen, I., Manecksha, R., McCullagh, E. & Al, E. (2012) The changing pattern of antimicrobial resistance within 42,033 *Escherichia coli* isolates from nosocomial, community, and urology patient-specific urinary tract infections, Dublin, 1999-2009. *BJU Int*, 109, pp. 1198- 1206.
- Dasgupta, P., Sacks, S., Khan, M. & Sheerin, N. (2005) 'Urinary Tract Infections; new insight into a common problem', *Postgrad. Med. J.*, pp. 8183-8186.
- Dejenie, S., Negeri, A. A., Legese, M. H., Bedada, T. L., Woldemariam, H. K. & Tullu, K. D. (2019) Extended-spectrum beta-lactamase production and multi-drug resistance among Enterobacteriaceae isolated in Addis Ababa, Ethiopia. *Antimicrobial Resistance and Infection Control* 8.

- George, A. & Levy, S. (1983) 'Amplifiable resistance to tetracycline, chloramphenicol, and other antibiotics in *Escherichia coli*: involvement of a non-plasmid-determined efflux of tetracycline', *J Bacteriol*, 155, pp. 531–540.
- Ghenghesh, K., Altomi, A., Gashout, S. & Abouhagar, B. (2003) 'High antimicrobial-resistance rates of *Escherichia coli* from urine specimens in Tripoli-Libya', *Garyounis Med J*, 20, pp. 89-93.
- Ghenghesh, K., Elkateb, E., Berbash, N., Nada, R., Ahmed, S., Rahouma, A. & Al, E. (2009) 'Uropathogens from diabetic patients in Libya: virulence factors and phylogenetic groups of isolated *Escherichia coli*', *J Med Microbiol.*, 58, pp. 1006_1014.
- Gupta, K., Hooton, T., Naber, K. & Al, E. (2011) 'International clinical practice guidelines for the treatment of acute uncomplicated cystitis and pyelonephritis in women: A 2010 update by the Infectious Diseases Society of America and the European Society for Microbiology and Infectious Diseases', *Clinical Infectious Diseases*, 52, pp. 1003-1012.
- Gupta, K., Hooton, T. & Stamm, W. (2001) 'Increasing antimicrobial resistance and the management of uncomplicated community-acquired urinary tract infections', *Annals of Internal Medicine*, 135, pp. 41-50.
- Hima-Lerible, H., Ménard, D. & Talarmin, A. (2003) 'Antimicrobial resistance among uropathogens that cause community-acquired urinary tract infections in Bangui, Central African Republic', *J Antimicrob Chemother*, 51, pp. 192-194.
- Hooton, T. (2003) 'Fluoroquinolones and resistance in the treatment of uncomplicated urinary tract infection', *Int. J. Antimicrob Agents*, 2, pp. 65-72.
- Hryniewicz, K., Szczy, K., Sulikowska, A., Jankowski, K., Betlejewska, K. & Hryniewicz, W. (2001) 'Antibiotic Susceptibility of bacterial strains isolated from urinary tract infections in Poland', *J. Antimicrob. Chemother*, 47, pp. 773-780.
- Institute of C. A. L. S. (2006) Performance standards for antimicrobial disk susceptibility tests. Wayne, PA USA: CLSI.
- Kahlmeter, G., Åhman, J. & Matuschek, E. (2015) 'Antimicrobial resistance of *Escherichia coli* causing uncomplicated urinary tract infections: a European update for 2014 and comparison with 2000 and 2008', *Infect Dis Ther*, 4, pp. 417–423.
- Kang, C., Kim, J., D.Wpark & Al, E. (2018) 'Clinical Practice Guidelines for the Antibiotic Treatment of Community-Acquired Urinary Tract Infections', *Infection & Chemotherapy*, 50, pp. 67–100.
- Karlowsky, J. A., Horban, D. J., Decorby, M. R., Lang, N. M. & Zhanel, G. G. (2006) 'Fluoroquinolone – Resistant Urinary Isolates of *E. coli* from outpatients Are Frequently Multidrug-Resistant: Results from the North American Urinary Tract Infection Collaborative Alliance – Quinolone Resistance Study', *Antimicrob. Agents Chemother*, 50, pp. 2251 -2254.
- Koen, B., Rahul, B., Amita, P., Jonathan, D., Julie, V. R. & Timo, S. (2017) 'Will co-trimoxazole resistance rates ever go down? Resistance rates remain high despite decades of reduced co-trimoxazole consumption', *J Glob Antimicrob Resist*, 11, pp. 71-74.
- Marei, A., Manzoor, A. & Mohammad, A. (2012) 'Extended-spectrum β -lactamase-producing *Escherichia coli* in clinical isolates in Benghazi, Libya: phenotypic detection and antimicrobial susceptibility pattern', *Medical Journal of Islamic World Academy of Science*, 20, pp. 49-56.
- Murray, P., Baron, E., Pfaller, M. & Tenover, F. (1999) Manual of Clinical Microbiology. Washington DC, American Society of Microbiology Press.
- Ndugulile, F., Jureen, R., Harthug, S., Urassa, W. & Langeland, N. (2005) Extended Spectrum β -Lactamases among Gram-negative bacteria of nosocomial origin from an Intensive Care Unit of a tertiary health facility in Tanzania. BMC Infect Dis 5.
- Obeng-Nkrumah, N., Twum-Danso, K., Krogfelt, K. & Newman, M. (2013) 'High levels of extended-Spectrum Beta-lactamases in a major teaching Hospital in Ghana: the need for regular monitoring and evaluation of antibiotic resistance', *Am J Trop Med Hyg.*, 89, pp. 960–964.
- Ouedraogo, A. S., Sanou, M., Kissou, A., Sanou, S., Solaré, H., Kaboré, F. & Al, E. (2016) High prevalence of extended-spectrum β -lactamase producing enterobacteriaceae among clinical isolates in Burkina Faso 2016;16(1):326. BMC Infect Dis., 16.
- Ozyurt, M., Oncul, O., Ceylan, S., Haznedaroglu, T. & Sahiner, F. (2008) 'Nurittin Ardic . Cross-resistance and associated resistance in *Escherichia coli* isolate from nosocomial urinary tract infections between 2004-2006 in a Turkish Hospital', *Cent. Eur. J. Med*, 3, pp. 446-452.
- Paterson, D. (2006) 'Resistance in Gram-negative bacteria: Enterobacteriaceae', *Am J infect control*, 34, pp. 20-28.
- Paterson, D., Mulazimoglu, L., Casellas, J. & Al, E. (2000) 'Epidemiology of ciprofloxacin resistance and its relationship to extended-spectrum beta-lactamase production in *Klebsiella pneumoniae* isolates causing bacteremia', *Clin Infect Dis*, 30, pp. 473–478.
- Sabrina, J., Said, A., Mabula, K., Eligius, F. & Samuel, Y. (2010) Antimicrobial resistance among producers and non-producers of extended-spectrum beta-lactamases in urinary isolates at a tertiary Hospital in Tanzania. BMC Research Notes, 3.
- Schwaber, M., Navon-Venezia, S., Schwartz, D., Schwaber, M., Navon-Venezia, S., Schwartz, D. & Al, E. (2005) 'High levels of antimicrobial Coresistance among extended-Spectrum-beta-lactamase-producing Enterobacteriaceae', *Antimicrob Agents Chemother.*, 49, pp. 2137–2139.
- Seitz, M., Stief, C. & Waidelich, R. (2017) Local epidemiology and resistance profiles in acute uncomplicated cystitis (AUC) in women: a prospective cohort study in an urban urological ambulatory setting. BMC Infect Dis, 16.
- Søråas, A., Sundsfjord, A., Sandven, I., Cathrine Brunborg, C., Jennum, P. (2013) Risk factors for community-acquired urinary tract infections caused by ESBL-producing enterobacteriaceae-a case-control study in a low prevalence country. PLoS One 23.
- Stapleton, P., Lundon, D., Mcwade, R., Scanlon, N., Hannan, M. M., O'Kelly, F., M Lynch, M. (2017) 'Antibiotic resistance patterns of *Escherichia coli* urinary isolates and comparison with antibiotic consumption data over 10 years, 2005-2014', *Ir J Med Sci*, 186, pp. 733–741.
- Tobgi, R., Taher, I. & Ali, M. (2001) 'Antibiotic susceptibility of uropathogens in Benghazi, Libya', *Jamahiriya Med J.*, 1, pp. 46-49.